

The effect of ecological disturbance on competition between Crinipellis perniciosa and other tropical fungi

Elizabeth Bravo-Velasquez and John Hedger

Department of Botany and Microbiology, The University College of Wales, Aberystwyth, Dyfed SY23 3DA, U.K.

Synopsis

es of the tropical fungus Crinipellis perniciosa obtained from cocoa and lianas in Ecuador were grown dia ranging from -0.45 MPa to -9.0 MPa water potential and their responses compared to those er agaries and Xylaria species isolated from cocoa. Antagonism between mycelia of C. perniciosa and isolates was also assessed over the same range of water potentials. It is hypothesised that niciosa and other canopy inhabiting fungi of tropical forests are adapted to water stress, but are not stitive with fungi which colonise later, following litterfall, from lower litter and soil horizons, where availability is higher. Xylaria spp. proved to be aggressive antagonists of C. perniciosa over a range ter potentials.

Introduction

ogists have emphasised the storied structure of tropical forests and the depenepiphytic communities. Hedger (1985) considered that species composition of l communities of saprotrophic agaric fungi is distinctly different from that of the r litter and soil horizons in tropical forest. He proposed that these agarics show tations to the daily and seasonal fluctuations in the water availability which r in the canopy of most tropical forests.

the study reported here we have used the Witches' Broom Disease fungus, pellis perniciosa (Stahel) Singer, as a possible example of a canopy-inhabiting c. The biology and ecology of this fungus, which occurs in Amazonia and ent tropical areas of South America, is reviewed elsewhere (Evans 1981; Rocha Hedger et al. 1987). It occurs both on cocoa (C-type) and on liana and other s in the forest (L-type). Infection of the canopy of cocoa by the basidiospores of be causes the development of large brooms in which, after death, a saprotrophic lium utilises the lignocellulose resource to produce flushes of basidiomes in wet ns. Optimum basidiome production has been shown by Rocha & Wheeler) to be dependent on daily fluctuations in the water content of the brooms. gard (1986), in researching the relationship between humidity and basidiome action by C. perniciosa in Rondonia, Brazil, found that brooms on the litter ce stayed wet for six hours a day longer than those in the canopy and produced icantly fewer basidiomes. Hedger (1985, 1988) also showed, in Ecuador, that ns incorporated into the litter beneath cocoa soon ceased to produce basidi-, and also found that the mycelium of C. perniciosa could not be recovered from ns buried for six months in the F1 layer of the litter. However, it persisted for up ee years in brooms suspended in the canopy.

ese observations indicate that the ecological disturbance, and associated change croclimatic conditions, of fall of brooms from canopy to litter may alter the

competitive balance between *C. perniciosa* and other broom-colonising fungi. In order to explore this possibility we have investigated interactions, over a range of water potentials, between the mycelium of *C. perniciosa* and other fungi isolated from witches' brooms. This model may represent the possible changes in water content which take place when brooms fall from the canopy into the litter layer.

Materials and methods

Isolation of fungi

Witches' brooms of cocoa were imported in the U.K. from Ecuador under licence and kept in cabinets under conditions similar to those described by Suarez (1977) and Rocha (1983), that is, 25–27°C with alternate 8 h, 16 h wet/dry cycles using a humidifier. Agarics which sporulated on the brooms were isolated by allowing basidiospores to deposit from basidiomes suspended over Potato Dextrose Agar (PDA) in petri dishes held at 25°C overnight. Resultant polyspore cultures were transferred to PDA slopes at 25°C. Cultures of *Xylaria* and *Nodulisporium gregarium* (Berk. & Curt.) Meyer were obtained by surface sterilising either 1.0 cm length broom sections, or 1.0 cm length ascocarp sections, for five minutes in 1% NaOCl solution, followed by washing in sterile distilled water, plating on PDA containing 0.05% Streptomycin at 25°C and subculturing from the margin of colonies after three to five days incubation. Cultures and dried material were deposited in the Herbarium, UCW Aberystwyth.

Response of mycelia to different water potentials

Isolates were grown on a basal medium of Glucose 2%, Malt Extract 2%, Peptone 0.1% and agar 1.5%, in 11 of distilled water. Potassium chloride was added to the medium before autoclaving to achieve a range of water potentials from -0.45 MPa to -13.8 MPa (Robinson & Stokes 1959). Growth rate was quantified in 9 cm petri dishes with medium inoculated centrally with 6 mm agar discs cut with a sterile cork borer from the margin of a colony, actively growing at 25°C on PDA agar. Mycelial extension was measured at 24 h intervals along three axes from the centre of the colony for up to one month. Petri dishes were kept in plastic bags, to prevent water loss.

Interactions at different water potentials were assessed by dual inoculation of plates with 6 mm discs placed 2.5 cm apart. Growth along three axes measured toward the confronting colony margin was compared with growth on the opposite side (Webber & Hedger 1985). In all cases, five replicate plates were prepared for each experimental condition.

Isolation of fungi

The following taxa were used for study:

Agaricales: Crinipellis perniciosa, Mycena theobromicola (Murr.) Dennis, Hohenbuehelia barbatula (Berk. & Curt.) Dennis, Clitopilus rhodotrama Sing., Crepidotus cuneiformis Pat., Coprinus jamaicensis Murr., Pluteus sp., Melanotus alpiniae (Berk.) Pilàt.

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Sphaeriales: *Xylaria grammica* (Mont.) Fr., *Xylaria multiplex* (Kze. ap. Fr.) Fr. omplex Berk. & Curt., *Xylaria* sp. 12, *Xylaria* sp. 59, *Xylaria* sp. 118.

Hyphomycetales: Nodulisporium gregarium.

All the taxa were isolated from carpophores on witches' brooms or plated out vitches' broom sections, with the exception of *Xylaria multiplex*, *X. grammica* and *Cylaria* 118, which were isolated from ascocarps growing on dead wood in cocoa itter. Each taxon was represented by only one isolate.

Results

Comparison of growth rates of isolates at different water potentials

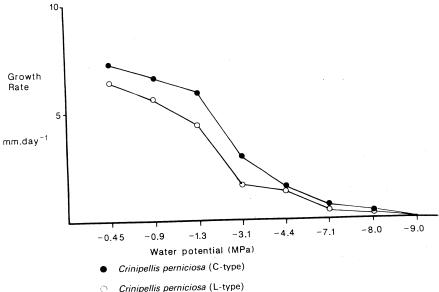
Cultures of C. perniciosa obtained from witches' brooms from Ecuador (C-type) roved to be tolerant of water stress, being able to develop small colonies at levels as ow as -8.0 MPa. Cultures obtained from basidiomes on liana (L-type), also from Ecuador, behaved similarly (see Fig. 1). Both grew optimally at -0.45 MPa. The even other agarics isolated from witches' brooms proved less tolerant of water stress and none grew at -4.4 MPa. Mycena theobromicola seemed particularly intolerant, with no growth detected at -3.1 MPa even after several months incubation at 25° C Fig. 1).

In contrast, isolates from *Xylaria*, or with a possible affinity as an anamorph in the ase of *Nodulisporium gregarium*, showed tolerance to water stress (Fig. 2). In some solates, growth at -0.9 MPa was equal to or slightly greater than at -0.45 MPa, and some growth was recorded at -9.0 MPa for most of the isolates, although, as with *C. perniciosa*, there was a long lag phase before growth occurred at this water potential.

The effects of water potential on interactions between isolates and C. perniciosa

The outcome of confrontation between C. perniciosa colonies and other agarics was learly influenced by the water potential of the medium. At -0.45 MPa in confrontations between other agarics and C. perniciosa, mycelial extension of both colonies ceased on contact, either accompanied by the formation of a pigment along he contact line (type 1 reaction) or by a clear zone between the two colonies (type 2 eaction). At lower water potentials (-0.9 MPa and -1.3 MPa), interactions changed from type 1 to type 2, with a decreased effect on the rate of C. perniciosa typhal growth on the side towards the opposing colony. Coprinus jamaicensis is an example (see Fig. 3, bottom). Except for C. jamaicensis, the isolates tested grew so slowly at -3.1 MPa that no contact occurred, and there was no effect on the C. perniciosa colony (Table 1).

In contrast, the *Xylaria* and *Nodulisporium gregarium* isolates proved to be iggressive invaders of C. perniciosa colonies over a range of water potentials (Table 2). Following contact, five of the isolates invaded the C. perniciosa colony more apidly at -1.3 MPa than at -0.45 and -0.9, although at -3.1 MPa and below here was a decrease in the amount of penetration of the C. perniciosa colony by the intagonists (see Table 2 and Fig. 3, top). *Xylaria grammica* was the only species which penetrated the C. perniciosa colony at -7.1 MPa.



Crinipellis perniciosa (L-type)

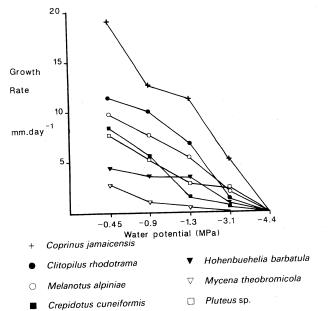


Figure 1. The effect of water potentials on the growth rates of isolates of C. perniciosa (C & L types) (upper figure) and seven other agarics (lower figure).

Typically the growth form of the mycelium of these antagonists changed on contact with the C. perniciosa colony. In most cases penetration of the colony was accompanied by vigorous formation of mycelial cords and fans, usually unpigmented, which were less developed in areas of the colony outside the sector occupying the C. perniciosa colony (Fig. 3, top). The cord forming sector often extended more quickly than the other parts of the colony, and rapidly covered the entire surface of

Figure 2

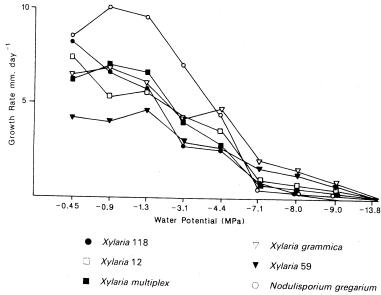
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- 2. The effect of water potentials on the growth rates of five isolates of Xylaria and Nodulisporium
- 1. Effect of water potential on interactions between Crinipellis perniciosa and other agarics isolated from witches' brooms.

	Water potential (MPa)					
Isolate	-0.45	-0.9	-1.3	-3.1	-4.4	
Coprinus jamaicensis	1	1	1	2	4	
Crepidotus cuneiformis	2	2	2	3	4	
Clitopilus rhodotrama	1	2	2	3	4	
Hohenbuehelia barbatula	1	2	2	3	4	
Melanotus alpiniae	1	1	2	3	4	
Mycena theobromicola	1	2	2	4	4	
Pluteus sp.	2	2	2	3	4	

Interaction Types (after one month's growth at 25°C):

- (1) deadlock, with formation of pigment at the confrontation line;
- (2) deadlock, with a clear zone of lysed mycelium at the confrontation line;
- (3) no contact;
- (4) no growth of antagonist.

perniciosa mycelium. In three isolates, at -0.45 MPa, however, only about the C. perniciosa colony area was occupied, followed by cessation of growth rple pigment formation in the contact zone. This variation in antagonism in 1 to the water potential of the mycelium is listed in Table 2.

Discussion

iciosa proved to be a water stress tolerant fungus, in comparison with the garics studied. In general, litter and wood colonising Basidiomycotina have

Table 2. The effect of water potential on interactions between *C. perniciosa* and *Xylaria* and *Nodulisporium*.

Organism	Water potential (MPa)							
	-0.45	-0.9	- i.3	-3.1	-4.4	-7.1		
Xvlaria 12	246	256	356	356	356	57		
Xylaria 59	346	346	346	346	34	7		
Xylaria 118	26	26	26	26	47	7		
X. grammica	346	36	36	3	3	1		
	346	346	346	36	36	7		
Nodulisporium	2	36	36	. 3	3	7		
X. multiplex	346	346	346	-	• •	7		

Interaction Types (after 1 month's growth at 25°C):

- (1) deadlock (no penetration of C. perniciosa colony);
- (2) partial invasion of *C. perniciosa* colony; (3) total invasion of *C. perniciosa* colony;
- (4) inhibition at distance of *C. perniciosa* growth before contact;
- (5) growth stimulation of test fungus mycelium before contact;
- (6) cord formation in *C. perniciosa* colony;
- (7) no contact.

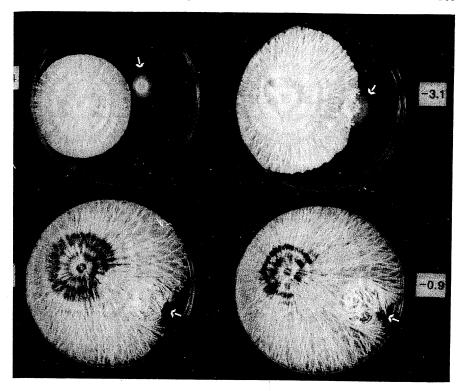
been found to be intolerant of low water potentials (Boddy 1983; Dix 1984; Dix & Frankland 1987), although studies have been confined to temperate species. In the case of C. perniciosa, the ability to continue growth at a low water potential must be an advantage in the seasonal and daily fluctuations in water stress in brooms in the cocoa canopy or in suspended liana debris. However, the alleviation of low water stress and the enrichment disturbance caused by the fall of brooms or liana debris into the litter layer changes the competitive balance between C. perniciosa and other fungi. Although only a limited selection of these fungi was evaluated as antagonists of C. perniciosa, the results of confrontation in agar over a range of water potentials show that the seven taxa were competitive only at high water potentials (-0.45 to -1.3 MPa, equivalent to high water contents in brooms and debris). At lower water potentials (less than -3.1 MPa, equivalent to dry conditions in debris or brooms) they made insufficient growth to be effective competitors. The mycelium of C. perniciosa can therefore retain lignocellulose resources in the brooms and liana debris in the canopy, but competition by other fungi is likely to increase after the "disturbance event" of the fall into the litter. Observations in the field confirmed that sporulation of the other agaries studied here such as Mycena theobromicola was most frequent on wet witches' brooms buried in cocoa litter, although they also occurred much less frequently on canopy brooms during wet periods (Hedger 1985).

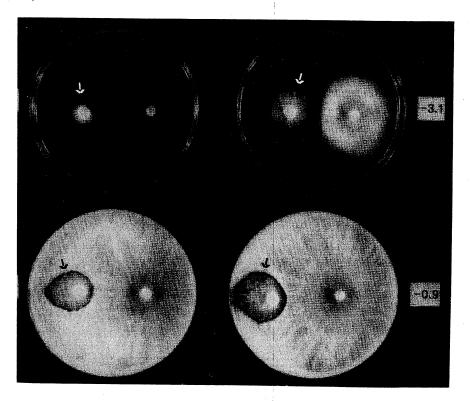
In contrast, the *Xylaria* species examined proved to be as tolerant as *C. perniciosa* of low water potentials and to be effective antagonists of this agaric even at -4.4 MPa. Rogers (1979), Boddy *et al.* (1985), and Whalley (1985) all consider that mycelia of temperate Xylariaceae are able to develop in dry conditions and to be more efficient as competitors at lower water potentials, a characteristic also present in the tropical species studied here. These fungi also have the potential to be canopy

Figure 3. Appearance of colonies of *C. perniciosa* and antagonists after three weeks' growth at 25°C on media with water potential -0.9, -1.3, -3.1 and -4.4 MPa; top: confrontation with *Xylaria* isolate 59; bottom: confrontation with *Coprinus jamaicensis*; position of the *C. perniciosa* inoculum is marked with an arrow.

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inhabiting and probably are able to compete for resources with C. perniciosa, and replace its mycelium, although the latter species is pre-established by an infective phase in witches' brooms.

It can be concluded that the disturbance represented by fall of litter from the canopy of tropical forest into the litter layer beneath marks a transition between dominance of resources by canopy inhabiting fungi, adapted to a water stressed environment, to occupation by communities of less stress adapted fungi.

Acknowledgments

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